Effect of Progenitor Cell Mobilization With Granulocyte-Macrophage Colony-Stimulating Factor in Patients With Peripheral Artery Disease: A Randomized Clinical Trial

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**IMPORTANCE** Many patients with peripheral artery disease (PAD) have walking impairment despite therapy. Experimental studies in animals demonstrate improved perfusion in ischemic hind limb after mobilization of bone marrow progenitor cells (PCs), but whether this is effective in patients with PAD is unknown.

**OBJECTIVE** To investigate whether therapy with granulocyte-macrophage colony-stimulating factor (GM-CSF) improves exercise capacity in patients with intermittent claudication.

**DESIGN, SETTING, AND PARTICIPANTS** In a phase 2 double-blind, placebo-controlled study, 159 patients (median [SD] age, 64 [8] years; 87% male, 37% with diabetes) with intermittent claudication were enrolled at medical centers affiliated with Emory University in Atlanta, Georgia, between January 2010 and July 2012.

**INTERVENTIONS** Participants were randomized (1:1) to received 4 weeks of subcutaneous injections of GM-CSF (leukine), 500 μg/day 3 times a week, or placebo. Both groups were encouraged to walk to claudication daily.

**MAIN OUTCOMES AND MEASURES** The primary outcome was peak treadmill walking time (PWT) at 3 months. Secondary outcomes were PWT at 6 months and changes in circulating PC levels, ankle brachial index (ABI), and walking impairment questionnaire (WIQ) and 36-item Short-Form Health Survey (SF-36) scores.

**RESULTS** Of the 159 patients randomized, 80 were assigned to the GM-CSF group. The mean (SD) PWT at 3 months increased in the GM-CSF group from 296 (151) seconds to 405 (248) seconds (mean change, 109 seconds [95% CI, 67 to 151]) and in the placebo group from 308 (161) seconds to 376 (182) seconds (change of 56 seconds [95% CI, 14 to 98]), but this difference was not significant (mean difference in change in PWT, 53 seconds [95% CI, −6 to 112], \( P = .08 \)). At 3 months, compared with placebo, GM-CSF improved the physical functioning subscore of the SF-36 questionnaire by 11.4 (95% CI, 6.7 to 16.1) vs 4.8 (95% CI, −0.1 to 9.6), with a mean difference in change for GM-CSF vs placebo of 7.5 (95% CI, 1.0 to 14.0; \( P = .03 \)). Similarly, the distance score of the WIQ improved by 12.5 (95% CI, 6.4 to 18.7) vs 4.8 (95% CI, −0.2 to 9.8) with GM-CSF compared with placebo (mean difference in change, 7.9 [95% CI, 0.2 to 15.7], \( P = .047 \)). There were no significant differences in the ABI, WIQ distance and speed scores, claudication onset time, or mental or physical component scores of the SF-36 between the groups.

**CONCLUSIONS AND RELEVANCE** Therapy with GM-CSF 3 times a week did not improve treadmill walking performance at the 3-month follow-up. The improvements in some secondary outcomes with GM-CSF suggest that it may warrant further study in patients with claudication.

**TRIAL REGISTRATION** clinicaltrials.gov Identifier: NCT01041417


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Peripheral artery disease (PAD) affects more than 8 million individuals in the United States.\textsuperscript{1-3} Although exercise, smoking cessation, antplatelet therapy, cilostazol, statins, and revascularization are used to treat PAD, men and women with PAD have significantly greater functional impairment and faster functional decline than those without PAD.\textsuperscript{1,4-7} Stem and progenitor cell (PC) functional impairment and faster functional decline than PAD, men and women with PAD have significantly greater 
cilostazol, statins, and revascularization are used to treat PAD holds.

Progenitor cells, particularly those of endothelial origin, are involved in vascular repair and regeneration.\textsuperscript{8} They originate primarily but not exclusively from the bone marrow, differentiate into endothelial and other vascular cells, and contribute to neovascularization during tissue repair by direct and paracrine mechanisms.\textsuperscript{8,9} Endogenous, pharmacologically stimulated, and endogenous PCs contribute to reendothelialization and neovascularization. Granulocyte colony-stimulating factor (G-CSF) and granulocyte-macrophage colony-stimulating factor (GM-CSF) stimulate mobilization of hematopoietic and other PCs from the bone marrow.\textsuperscript{10,11} In the murine hind limb ischemia model, GM-CSF administered by injection or by plasmid transfer augments circulating levels of PCs, increases capillary density, and promotes arteriogenesis.\textsuperscript{12} Granulocyte-macrophage colony-stimulating factor also augments neoendothelialization of denuded arteries and promotes proliferation, differentiation, and survival of hematopoietic cells, monocytes, and macrophages.\textsuperscript{9,10,13,14}

Based on the observations of improved neovascularization in experimental models with GM-CSF,\textsuperscript{9} and some equivocal clinical trials,\textsuperscript{15-17} we previously completed a phase I dose-escalation trial in patients with claudication, which demonstrated the safety of GM-CSF and its ability to mobilize PCs into the circulation. Additionally, we observed a strong trend toward an improvement in exercise duration after 3 months of treatment.\textsuperscript{18} In this larger phase 2 study, we tested the hypothesis that GM-CSF administration in patients with symptomatic PAD would improve walking performance. In exploratory analyses, we assessed the safety of GM-CSF for patients with claudication.

Methods

Between January 2010, and July 2012, 322 patients aged between 21 and 80 years with PAD were screened. Of these, 159 were enrolled from Emory Healthcare, Veterans Affairs hospitals in Atlanta, and the Medical College of Georgia (Figure). Practicing physicians at these hospitals were invited to refer potentially eligible patients with PAD.

Eligibility criteria required participants to have angiographically documented history of obstructive PAD, a 2-month or longer history of intermittent claudication (Rutherford category 1-3)\textsuperscript{19} in one or both lower extremities despite appropriate and stable medication regimen that included a statin for at least 3 months, a peak walking time (PWT) on a treadmill between 1 and 12 minutes, and an ankle brachial index (ABI) of less than 0.85 in the symptomatic limb. At screening, participants had to exhibit stable and reproducible walking capacity, defined as a less than 20% variation in PWT measured by 2 screening treadmill tests within a 2-week period. If the ABI was greater than 0.85, a reduction of 20% in ABI measured within 1 minute of treadmill testing or a toe brachial index of less than 0.70 was required for eligibility.

Exclusion criteria included the presence of critical limb ischemia (Rutherford category 4-6); advanced diabetic retinopathy; current or planned participation in a structured exercise program; active infection; coronary or lower extremity revascularization within the past 4 months; history of acute coronary or cerebrovascular syndrome within the past 4 months; any history of myeloid malignancy, severe congestive heart failure, or chronic renal, hepatic, or other inflammatory diseases; or a life expectancy of less than 12 months. The trial was
approved by the institutional review boards, informed consent was obtained from participants, and an investigational new drug waiver was obtained from the US Food and Drug Administration.

**Randomization and Study Interventions**

Treatment assignments were generated in random, permuted blocks by the study pharmacy at Emory and stratified for diabetes to ensure equal randomization of participants with diabetes into the treatment groups. Participants were randomized 1:1 to receive either GM-CSF, 500 μg (sargramostim [Leukine]; sanofi-aventis), or matching placebo (normal saline) by a self-administered subcutaneous injection thrice weekly on Monday, Wednesday, and Friday for 4 weeks. At the beginning of each week, 1 injection was administered under observation. Patients were advised to walk to claudication only at least 3 times a day during the 4-week treatment period to encourage “homing” of the mobilized PCs to the ischemic muscle bed in the GM-CSF group. The dose of study drug was modified by an investigator who was not involved directly with patients (E.K.W.) for adverse effects, including pain unresponsive to analgesics, rash on more than 25% of body surface area, fever greater than 38.5°C, splenomegaly, or significant leukocytosis (white blood cell count >35 000/μL) or thrombocytopenia (platelet count <75 000/μL). Participants and investigators involved in the clinical evaluation, implementation of the protocol, and analysis remained blinded to treatment assignment for the duration of the trial.

**Measurements**

During weekly visits at the end of each of the first 4 weeks, adherence was reinforced. Safety and adverse events were recorded at each visit (baseline; weeks 1, 2, 3, 4, and 5; and 3 and 6 months) and by telephone interview at 8 weeks. Measures of response—treadmill exercise, ABI, and symptom questionnaires—were recorded at baseline and 3 and 6 months. Measures of underlying mechanisms (PC counts) were recorded at baseline, 2 weeks, 4 weeks, 3 months, and 6 months. (Details on this timeline appear in the eFigure in the Supplement.)

**Treadmill Testing**

Treadmill exercise testing was conducted using the protocol by Gardner et al.20 The modified Bruce protocol was used among those participants (n = 6) who developed claudication after exercising for 12 minutes on the Gardner et al protocol. Treadmill exercise testing was performed twice at each visit, with the maximum claudication onset time (COT) and PWT recorded. The COT was measured as the time to onset of the participant’s typical claudication and the PWT as the maximum distance the patient could walk on the treadmill. Patients unfamiliar with treadmill exercise were allowed to familiarize themselves with the treadmill beforehand. Patients underwent 2 treadmill exercise tests to ensure reproducibility (<20% difference in PWT) and a third exercise test for cardiorespiratory testing (Spectra 29; SensorMedics) to measure peak oxygen uptake. During the 3- and 6-month visits, patients underwent 2 exercise tests within a 1-week period with cardiopulmonary testing during one test. The maximum COT and PWT from the tests was used for data analysis at each time point.

**Ankle Brachial Index**

With the patient lying supine in a temperature-controlled room, blood pressure cuffs were placed bilaterally on the upper arms and ankles and inflated to 30 mm Hg above the systolic pressure (Flo-Laboratory Model 2100; Parks Medical Electronics). Blood pressure in bilateral brachial, posterior tibial, and dorsalis pedis arteries was measured by Doppler. The ABI was calculated as the ratio of the higher of the 2 ankle pressures to the higher of the 2 brachial pressures.

**Questionnaires**

Participants completed the walking impairment questionnaire (WIQ), which enabled calculation of walking distance, walking speed, and stair-climbing scores. The 36-item Short-Form Health Survey (SF-36) was completed, and the physical composite score (PCS-36), mental composite score (MCS-36), and physical functioning subscore were calculated.

**Progenitor Cell Counts**

After an overnight fast, venous blood was collected in EDTA tubes and incubated with fluorochrome-labeled monoclonal antihuman mouse antibodies within 4 hours. Cell populations enriched for circulating PCs were enumerated using flow cytometry as CD45med+ cells coexpressing CD34+, CD133+, vascular endothelial growth factor receptor 2 (VEGFR2+), or CXCR4+ and their combination.21-24

We incubated 300 μL of peripheral blood with 7 μL of FITC-CD34 (BD Biosciences), PerCP-CD45 (BD Biosciences), PE-VEGF-R2 (R&D System) and 5-μL APC-CD133 (Miltenyi), and 3-μL PE-Cy7-conjugated anti-CXCR4 (EBioscience, clone 12G5) in the dark for 15 minutes. Then 1.5-mL ammonium chloride lysing buffer was added to lyse red blood cells; 1.5-mL staining medium (PBS with 3% heat-inactivated serum and 0.1% sodium azide) was added to stop the lysing reaction. Prior to flow cytometry, 100 μL of AccuCheck Counting Beads (Invitrogen, No. PC100) were added to act as an internal standard for direct estimation of the concentration of target cell subsets. At least 2.5 million events were acquired from the cytometer. Flow data were analyzed with Flowjo software (TreeStar). Absolute mononuclear cell count was estimated as the sum of lymphocytes and monocytes using a Coulter ACT/Diff cell counter (Beckman Coulter). Progenitor cell populations are reported as cell counts per μL. In 20 samples that were analyzed on 2 occasions by the same technician, the coefficients of variation of the cell types were as follows: CD34+, 2.9%; CD34+/CD133+, 4.8%; CD34+/CXCR4+, 6.5%; and CD34+/CD133+/CXCR4+, 7.5%, CD34+/VEGFR2 cells, 21.6%.

**Statistical Analysis**

The primary efficacy outcome was change in PWT from baseline at 3 months. Secondary efficacy outcomes considered were change in PWT at 6 months, change in COT at 3 and 6 months, and change in the WIQ and SF-36 survey scores. Safety end points included frequencies of adverse events, serious ad-
verse events, and discontinuations due to adverse events. All participants who were randomized and had end-point measurements at 3 or 6 months were included in the efficacy and safety analyses on an intention-to-treat basis. A post hoc per-protocol analysis was also performed after excluding protocol deviations.

Continuous variables are presented as mean (SD). Changes from baseline at 3 and 6 months are presented as mean (95% CI). Normally distributed data are presented as median (interquartile range [IQR]) and categorical variables as proportions. Group comparisons involving categorical and continuous outcomes at baseline were evaluated with χ² tests and t tests, respectively. Statistical analyses of the primary and secondary outcomes were performed using a linear mixed-effects model, which provided separate estimates of the mean change by time on intervention and control groups. The model-based means are unbiased with unbalanced and incomplete data, provided that the missing data are noninformative (missing at random). A compound symmetry form in the repeated measurements was assumed for each outcome. Safety end points were analyzed by χ² tests. Data analysis was performed using SAS version 9.3 (SAS Institute). Statistical significance was based on 2-tailed tests, and a significance level of α = .05 was used. Based on our phase 1 study, a target sample size of 80 patients in each group allowed us to detect a minimum difference in the primary endpoint, comparing the increase in PWT from baseline to 3 months between the GM-CSF group (from mean [SD] 296 [151] to 405 [248] seconds, a change of 109 seconds [95% CI, 67 to 151], 30% increase) and placebo group (from 308 [161] to 376 [182] seconds, a mean change of 68 seconds [95% CI, 14 to 98], 21% increase), was not significant (difference in change between groups of 53 seconds [95% CI, −6 to 112]; P = .08) (Table 3). Changes in PWT at 6 months and COT at both 3 and 6 months were also not significant.

Effect of GM-CSF on Other Secondary End Points
There was a greater improvement in the walking distance sub-score of the WIQ at 3 months with GM-CSF compared with placebo; the difference in change between groups was 7.9 (95% CI, 0.2-15.7; P = .047) (Table 3). Similarly, there was a significantly greater improvement in the physical functioning subscale score of the SF-36 survey at 3 months with GM-CSF compared with placebo; the difference in change between groups was 7.5 (95% CI, 1.0-14.0; P = .03) (Table 3). There were no significant differences in changes in the WIQ speed or stair-climbing scores between the GM-CSF and placebo groups. There were no significant differences in changes in the PCS-36 or MCS-36 scores between the GM-CSF and placebo groups. In addition, there were no significant differences in changes in the ABI or in oxygen consumption at the 3- or 6-month follow-up visits between the 2 groups.

Adverse Effects
Adjusting the GM-CSF dose was necessary for 1 patient who had a platelet count less than 75 000. There were 18 serious adverse events, 9 in the placebo and 9 in the GM-CSF group (eTable 1 in the Supplement). One patient in the GM-CSF group died in an automobile crash. Cancer was diagnosed in 2 patients treated with GM-CSF and 1 patient treated with placebo. Six of these patients were not followed up after their serious adverse events (Figure). Nonserious adverse effects that were more frequently observed in the GM-CSF group compared with the placebo group included headache, gastrointestinal symptoms, rash, and fatigue and were restricted to the first 4 weeks of therapy (eTable 2 in the Supplement). The most common adverse effect was rash appearing at the injection site, but none of these led to discontinuation or modification of therapy.

Exploratory Analyses
After excluding 4 participants with protocol deviations, including incomplete treatment due to incarceration, acci-
In an exploratory analysis, we also investigated the relationship between the magnitude of PC mobilization and the improvement in exercise capacity with GM-CSF. Participants receiving GM-CSF who had a greater increase in PCs (>100% increase in CD34+/CD133+ cells, n = 38) at 2 weeks had a significantly greater increase in PWT at 3 months (131 seconds [95% CI, 74-187] vs 60 seconds [95% CI, 26-93], P = .04) and at 6 months (136 seconds [95% CI, 80-192] vs 70 seconds [95% CI, 37-103], P = .048) than those with less than 100% increase in these PCs.
Table 2. Changes in Leucocyte Subsets With GM-CSF and Placebo

<table>
<thead>
<tr>
<th>Leukocytes, ×10^6 cells/mL</th>
<th>Mean (95% CI)</th>
<th>P Value*</th>
</tr>
</thead>
<tbody>
<tr>
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<tr>
<td>GM-CSF</td>
<td></td>
<td></td>
</tr>
<tr>
<td></td>
<td>7.2</td>
<td>8.4</td>
</tr>
<tr>
<td></td>
<td>(6.8 to 7.7)</td>
<td>(7.7 to 9.0)</td>
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<td>(6.7 to 7.6)</td>
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<tr>
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<td>(0.9 to 2.2)</td>
<td>(2.2 to 3.5)</td>
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<tr>
<td>CD34&lt;sup&gt;+&lt;/sup&gt;, cells/μL</td>
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<tr>
<td></td>
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<td>(4.1 to 5.7)&lt;sup&gt;b&lt;/sup&gt;</td>
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<tr>
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<tr>
<td></td>
<td>(1.9 to 2.6)</td>
<td>(1.8 to 2.3)</td>
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<tr>
<td>Difference&lt;sup&gt;c&lt;/sup&gt;</td>
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<td>(2.2 to 3.5)</td>
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<tr>
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<tr>
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<td>2.5</td>
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<td></td>
<td>(1.0 to 1.3)</td>
<td>(2.0 to 3.0)&lt;sup&gt;b&lt;/sup&gt;</td>
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<tr>
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<td>1.0</td>
</tr>
<tr>
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<td>(1.0 to 1.3)</td>
<td>(0.9 to 1.1)</td>
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<tr>
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<td>(1.1 to 1.7)</td>
<td>(0.9 to 1.1)</td>
</tr>
<tr>
<td>CD34&lt;sup&gt;+&lt;/sup&gt;/CXCR4&lt;sup&gt;+&lt;/sup&gt;, cells/μL</td>
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<tr>
<td>GM-CSF</td>
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<td>2.5</td>
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<td>(2.0 to 3.0)&lt;sup&gt;b&lt;/sup&gt;</td>
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<td>(0.9 to 1.2)</td>
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<td>0.5</td>
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<tr>
<td></td>
<td>(1.0 to 1.7)</td>
<td>(0.4 to 0.6)</td>
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Abbreviation: GM-CSF, granulocyte-macrophage colony-stimulating factor.  
* P values refer to differences between GM-CSF and placebo groups by mixed-effects models.  
<sup>b</sup> P < .001 from t tests comparing baseline at week 2.  
<sup>c</sup> Difference of changes from baseline between GM-CSF and placebo.

Discussion

In this phase 2 study, we tested the hypothesis that administration of a myeloid cytokine that mobilizes bone marrow PCs into the circulation would improve walking performance in patients with claudication. In a placebo-controlled, double-blind study of subcutaneous injections of GM-CSF (500 μg per day self-administered 3 times a week for 4 weeks), there was no difference in change in PWT between the GM-CSF and placebo groups. In secondary analyses, there was improvement in the SF-36 physical functioning score and the WIQ distance score with GM-CSF as compared with placebo. However, there were no significant differences between the intervention and placebo groups with regard to change in WIQ walking speed or stair-climbing score, COT, PCS-36 or MCS-36 components of the SF-36, ABI, or oxygen consumption.

The crucial contribution of PCs to vascular repair, collateralization, and re-endothelialization of the vasculature after acute ischemic injury is well established. Although GM-CSF is a less potent mobilizing agent for leukocytes and hematopoietic progenitors compared with G-CSF, potential advantages of GM-CSF include selective mobilization of more primitive stem cells (CD34+ /CD38− /HLA-DR+) and macrophages. Moreover, GM-CSF has direct stimulatory effects on endothelial cells and monocytes, which may contribute to its vasculogenic properties. Mobilization of the bone marrow cells is relatively modest with GM-CSF, addressing potential safety concerns that have been raised with the use of G-CSF in atherosclerosis. There may be several reasons for the lack of improvement in our primary outcome and most of our secondary outcome measures in this PAD population. First, aging and exposure to cardiovascular risk factors that are prevalent in our population are known to reduce the number and potency of endogenous PCs. The increase in leukocyte counts in our PAD population was modest in comparison with healthy cohorts, probably a reflection of senescence of enhanced mobilization in those with relatively modest issues with the self-injectable treatment in our study may have reduced effectiveness in some participants. Study drugs were injected under observation once each week. Whether titration to a higher dose of GM-CSF would have enhanced mobilization in those with relatively modest mobilization remains to be studied. However, in our phase 1 dose-escalation study, we did not observe a dose-response in the magnitude of PC mobilization with GM-CSF given in concentrations ranging from 3 to 10 μg/kg per day. Thus, it is possible that some participants were unable to mobilize sufficient PCs from their marrow for cytokine therapy to be effective or that stronger cytokines such as G-CSF may be appropriate for these individuals.
We observed a modest improvement in PWT in our phase 1 study after 2 weeks of therapy with GM-CSF.18 In this phase 2 study, we increased the duration of treatment to 4 weeks and selected a 500-μg dose of GM-CSF that was equivalent to a dose of approximately 6 μg/kg for an 80-kg individual. Leukocyte count data demonstrated that peak mobilization of bone marrow cells occurred at 3 weeks with a decline toward baseline count data demonstrated that peak mobilization of bone marrow of approximately 6 μg/kg for an 80-kg individual. Leukocyte selected a 500-μgdose of GM-CSF that was equivalent to a dose 2 study, we increased the duration of treatment to 4 weeks and  

Table 3. Peak Walking Time, Claudication Onset Time, ABI, Peak Oxygen Consumption, WIQ and SF-36 Scores, and Their Changes at 3 and 6 Months With GM-CSF and Placebo*

<table>
<thead>
<tr>
<th></th>
<th>GM-CSF</th>
<th>Placebo</th>
<th>Difference Between GM-CSF and Placebo</th>
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<tr>
<td></td>
<td>Baseline (s)</td>
<td>3 mo (s)</td>
<td>6 mo (s)</td>
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<tr>
<td>PWT, s</td>
<td>296 (151)</td>
<td>405 (248)</td>
<td>411 (253)</td>
</tr>
<tr>
<td>Change</td>
<td>0.19 (0.06)</td>
<td>0.20 (0.06)</td>
<td>0.20 (0.06)</td>
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<tr>
<td>COT, s</td>
<td>146 (67)</td>
<td>230 (161)</td>
<td>233 (158)</td>
</tr>
<tr>
<td>Change</td>
<td>0.05 (0.04)</td>
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<tr>
<td>ABI</td>
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<td>0.63 (0.02)</td>
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<tr>
<td>V̇O₂max, L/min</td>
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<td>1.24 (0.05)</td>
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<tr>
<td>SF-36 d</td>
<td>40.2 (27.6)</td>
<td>34.5 (27.4)</td>
<td>29.7 (22.7)</td>
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<td>PCS-36</td>
<td>31.1 (7.9)</td>
<td>35.5 (8.2)</td>
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<tr>
<td>Physical functioning</td>
<td>50.1 (19.1)</td>
<td>52.8 (20.6)</td>
<td>57.4 (25.9)</td>
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<td>Change</td>
<td>1.4 (1.3)</td>
<td>0.8 (1.3)</td>
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</table>

Abbreviations: ABI, ankle brachial index; COT, claudication onset time; GM-CSF, granulocyte-macrophage colony-stimulating factor; MCS-36, mental composite score; PSC-36, physical composite score; PWT, peak walking time; SF-36, 36-item Short-Form Health Survey; WIQ, walking impairment questionnaire.

* Data are presented as absolute mean (standard deviation). For change from baseline, and for differences between groups, data are presented as model-based means (and 95% confidence intervals).

P values refer to differences between GM-CSF and placebo groups by mixed-effects models at 3 months and 6 months.

The WIQ quantifies walking difficulty on a 100-point scale, in which 0 indicates extreme difficulty and 100 indicates no difficulty with walking distance, walking speed, and stair-climbing elements.

The SF-36 PCS-36 and MCS-36 are summary measures derived from 8 scale scores, including the physical functioning subscale. Higher scores indicate better performance.
tion consisted of patients with claudication who underwent angiography. Further study is needed to determine whether GM-CSF is beneficial in critical limb ischemia or to other people not eligible for this study. Third, our study design encouraged participants in both the GM-CSF and placebo groups to walk to claudication several times a day, to promote homing of PCs to the ischemic muscles. However, this increased activity may have resulted in greater improvement in walking capacity in participants receiving placebo, as expected from studies demonstrating improvements with structured exercise in PAD. Fourth, the magnitude of change in PWT that we used to represent a clinically meaningful change. The magnitude of improvement in PWT from GM-CSF was substantially less than that previously reported for ramipril therapy. Fifth, although we had based our sample size for this study on our phase 1 findings, we observed greater variability in PWT, and thus this study may have been underpowered for our primary end point. Sixth, although no significant differences were observed in the serious adverse event rates between groups, our study was not powered to evaluate this.

Conclusions

Therapy with GM-CSF 3 times a week did not improve treadmill walking performance at 3-month follow-up. The improvement in a subset of secondary outcomes observed with GM-CSF suggests that GM-CSF may warrant further study in patients with claudication. In addition, further investigation is needed to investigate the variability of responsiveness to GM-CSF and its clinical significance.

ARTICLE INFORMATION

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